

## **BAB V**

### **KESIMPULAN DAN SARAN**

#### **A. Kesimpulan**

Berdasarkan penelitian yang telah dilakukan maka dapat diambil kesimpulan bahwa :

Pertama, bakteri *Escherichia coli* ATCC 25922, *Pseudomonas aeruginosa* ATCC 27853, *Salmonella typhi* ATCC 13311, *Serratia marcescens* ATCC 8100, dan *Staphylococcus aureus* ATCC 25923 menghasilkan enzim amilase.

Kedua, dari kelima amilase bakteri yang memiliki aktivitas enzim amilase paling besar adalah *Escherichia coli* ATCC 25922 dengan diameter zona bening sebesar 19,79 mm.

Ketiga, karakteristik *Escherichia coli* ATCC 25922 memiliki aktivitas paling besar pada suhu 30°C dan pH 8. *Pseudomonas aeruginosa* ATCC 27853 memiliki aktivitas paling besar pada suhu 50°C dan pH 8. *Salmonella typhi* ATCC 13311 memiliki aktivitas paling besar pada suhu 50°C dan pH 8. *Serratia marcescens* ATCC 8100 memiliki aktivitas paling besar pada suhu 50°C dan pH 8. *Staphylococcus aureus* ATCC 25923 memiliki aktivitas paling besar pada suhu 50°C dan pH 8.

#### **B. Saran**

Perlu dilakukan penelitian lanjutan untuk sampai pada tahap pemurnian enzim amilase dan pengukuran kadar protein yang terkandung, karena yang dilakukan dalam penelitian ini masih berupa ekstrak kasar amilase saja. Dan juga pada penelitian aktivitas enzim harus lebih teliti dan memahami faktor apa saja yang mempengaruhi aktivitas enzim selain pH, suhu, dan substrat.

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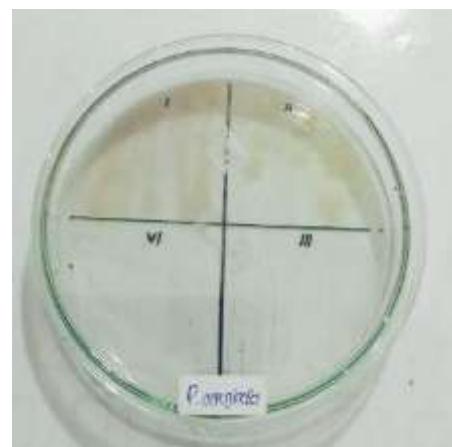
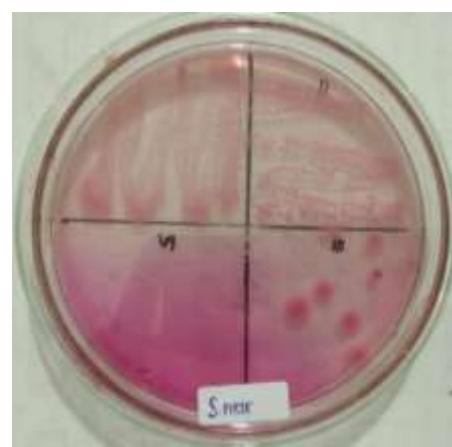
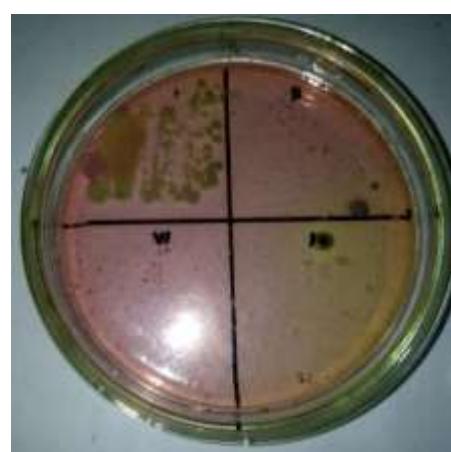
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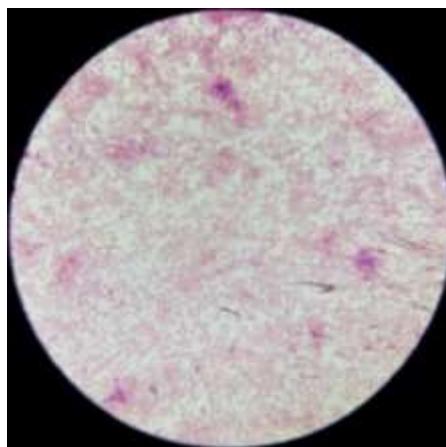
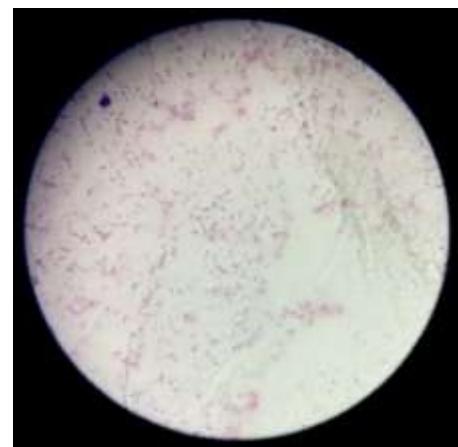
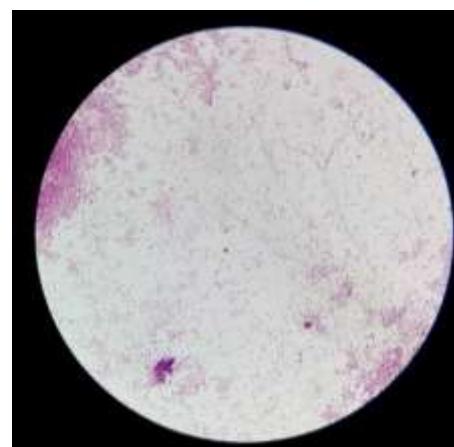
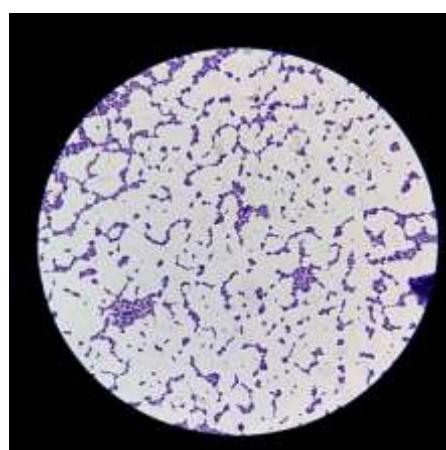
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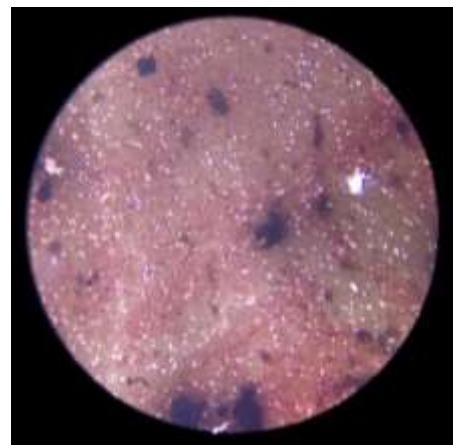
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**Lampiran 1. Hasil Identifikasi Bakteri berdasarkan Koloni**Koloni *E. coli*Koloni *P. aeruginosa*Koloni *S. typhi*Koloni *S. marcescens*Koloni *S. aureus*

**Lampiran 2. Hasil Pewarnaan Gram Bakteri***E. coli**P. aeruginosa**S. typhi**S. marcescens**S. aureus*

**Lampiran 3. Hasil Pewarnaan Kapsul***P. aeruginosa**S. typhi*

**Lampiran 4. Hasil Uji Biokimia**

Bakteri	SIM	KIA	LIA	SCA
<i>E. coli</i>				
<i>P. aeruginosa</i>				
<i>S. typhi</i>				
<i>S. marcescens</i>				

**Lampiran 5. Hasil Uji Katalase dan Uji Koagulase *S. aureus***

Uji Katalase



Uji Koagulase

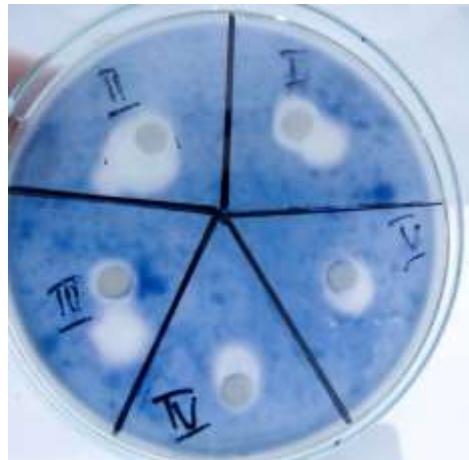
**Lampiran 6. Produksi Enzim Amilase**

Fermentasi bakteri pada media produksi enzim



Ekstrak kasar enzim amilase setelah disentrifugasi

**Lampiran 7. Hasil Zona Bening Amilase Bakteri**

Replikasi	Zona Bening
1	 A Petri dish containing a blue agar medium with five distinct bacterial spots. The dish is divided into four quadrants by black lines, and each quadrant is further divided into two sectors by diagonal lines, creating a total of eight angular sectors. The sectors are labeled with Roman numerals: I (top-right quadrant, top sector), II (top-right quadrant, bottom sector), III (bottom-left quadrant, left sector), IV (bottom-left quadrant, right sector), V (top-left quadrant, top sector), VI (top-left quadrant, bottom sector), VII (bottom-right quadrant, left sector), and VIII (bottom-right quadrant, right sector). Zone I shows a large, well-defined clear zone around the spot. Zones II, III, and IV show smaller, partially confluent clear zones. Zone V shows a very small or no discernible zone. Zone VI is not clearly visible. Zone VII shows a small clear zone. Zone VIII shows a small clear zone.
2	 A Petri dish containing a blue agar medium with five distinct bacterial spots. The dish is divided into four quadrants by black lines, and each quadrant is further divided into two sectors by diagonal lines, creating a total of eight angular sectors. The sectors are labeled with Roman numerals: I (top-right quadrant, top sector), II (top-right quadrant, bottom sector), III (bottom-left quadrant, left sector), IV (bottom-left quadrant, right sector), V (top-left quadrant, top sector), VI (top-left quadrant, bottom sector), VII (bottom-right quadrant, left sector), and VIII (bottom-right quadrant, right sector). Zone I shows a large, well-defined clear zone. Zones II, III, and IV show smaller, partially confluent clear zones. Zone V shows a very small or no discernible zone. Zone VI is not clearly visible. Zone VII shows a small clear zone. Zone VIII shows a small clear zone.
3	 A Petri dish containing a blue agar medium with five distinct bacterial spots. The dish is divided into four quadrants by black lines, and each quadrant is further divided into two sectors by diagonal lines, creating a total of eight angular sectors. The sectors are labeled with Roman numerals: I (top-right quadrant, top sector), II (top-right quadrant, bottom sector), III (bottom-left quadrant, left sector), IV (bottom-left quadrant, right sector), V (top-left quadrant, top sector), VI (top-left quadrant, bottom sector), VII (bottom-right quadrant, left sector), and VIII (bottom-right quadrant, right sector). Zone I shows a large, well-defined clear zone. Zones II, III, and IV show smaller, partially confluent clear zones. Zone V shows a very small or no discernible zone. Zone VI is not clearly visible. Zone VII shows a small clear zone. Zone VIII shows a small clear zone.

**Lampiran 8. Larutan Buffer**

Pengukuran pH

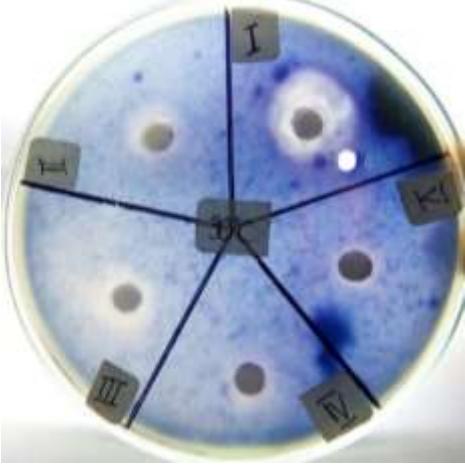
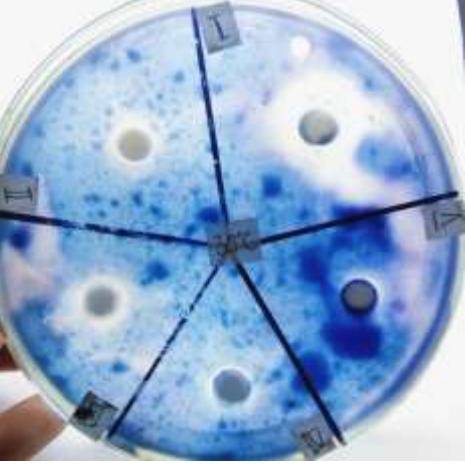


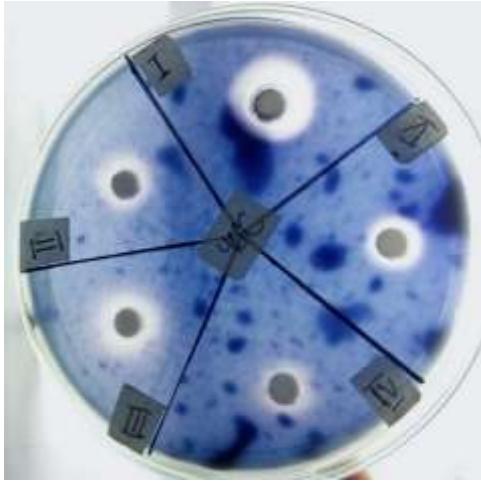
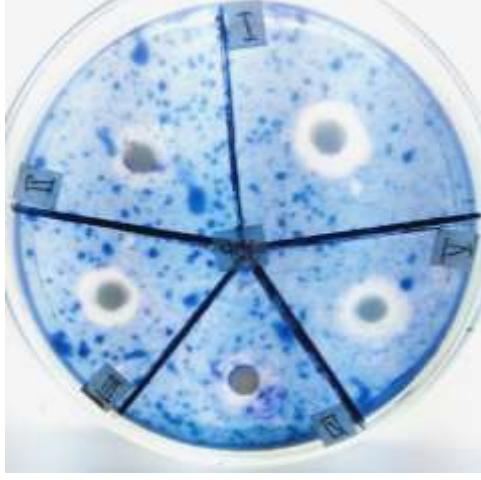
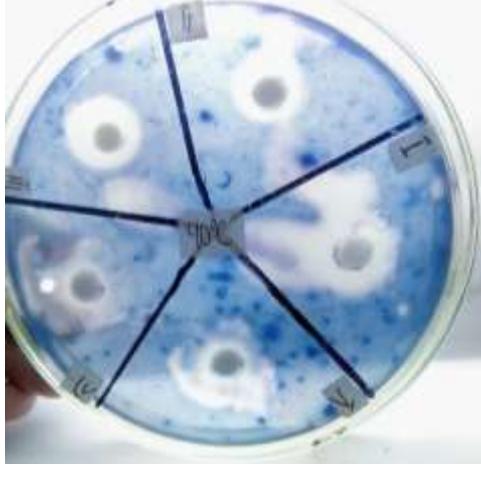
Larutan buffer sitrat (pH 4-6) dan buffer fosfat (pH 7-8)

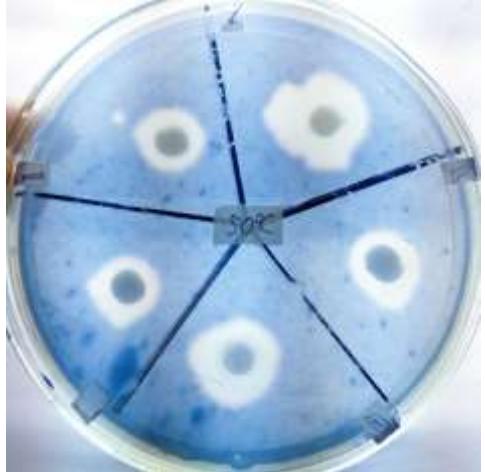
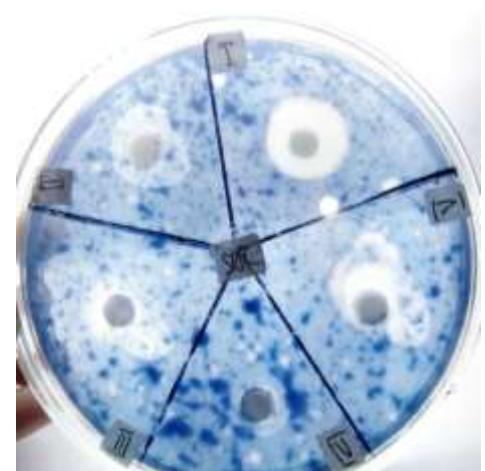
**Lampiran 9. Ekstrak Kasar Amilase Variasi Suhu**

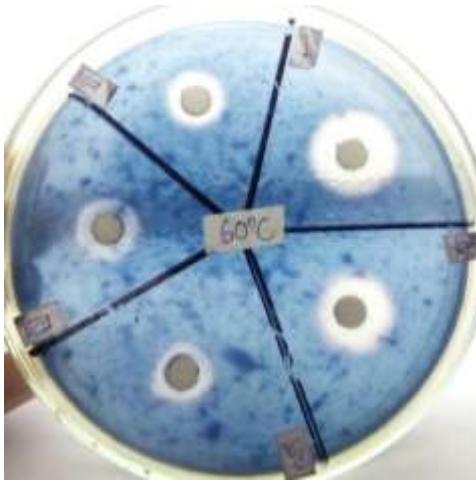
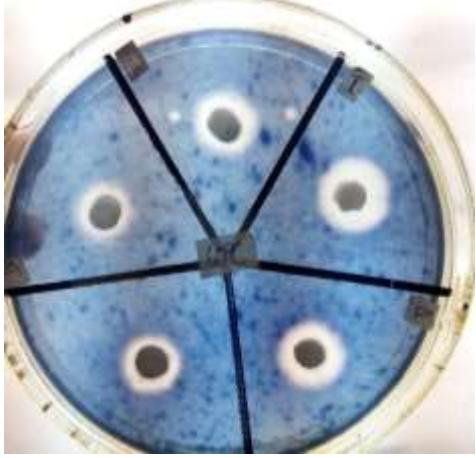
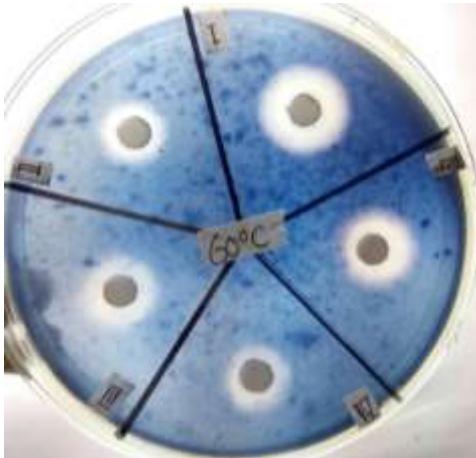
**Lampiran 10. Ekstrak Kasar Amilase Variasi pH**

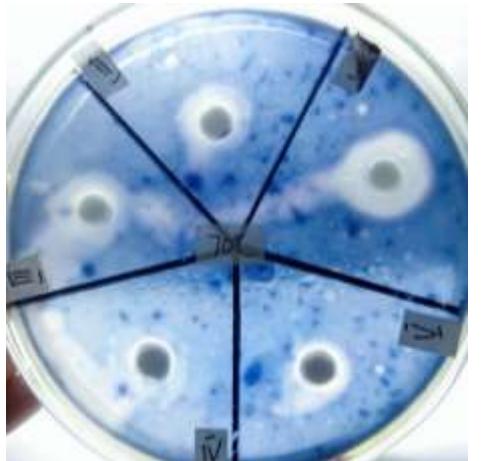
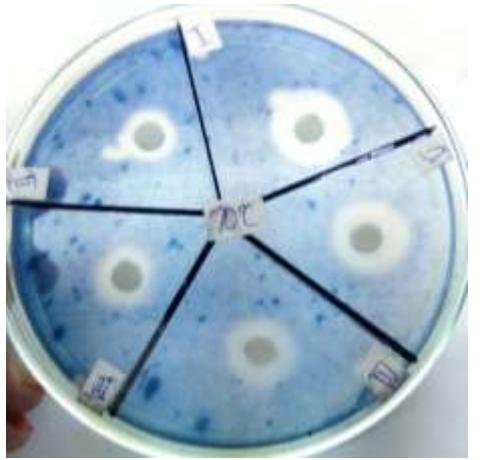
**Lampiran 11. Zona Bening Aktivitas Amilase terhadap Suhu**

Replikasi	Zona Bening
1	
2	
3	

Replikasi	Zona Bening
1	 A petri dish with a blue agar medium. Four dark grey rectangular discs are placed at the center and the corners of a square grid drawn on the dish. Four distinct clear zones of inhibition (benign zones) are visible, each centered on one of the discs, labeled I, II, III, and IV.
2	 A petri dish with a blue agar medium. Four dark grey rectangular discs are placed at the center and the corners of a square grid drawn on the dish. Four distinct clear zones of inhibition (benign zones) are visible, each centered on one of the discs, labeled I, II, III, and IV.
3	 A petri dish with a blue agar medium. Four dark grey rectangular discs are placed at the center and the corners of a square grid drawn on the dish. Four distinct clear zones of inhibition (benign zones) are visible, each centered on one of the discs, labeled I, II, III, and IV.

Replikasi	Zona Bening
1	 A petri dish containing a blue agar medium with four distinct white bacterial growth inhibition zones arranged in a cross pattern. The zone in quadrant I is the largest and most prominent.
2	 A petri dish containing a blue agar medium with four distinct white bacterial growth inhibition zones arranged in a cross pattern. The zones in quadrants I and II are larger and more confluent than those in the other quadrants.
3	 A petri dish containing a blue agar medium with four distinct white bacterial growth inhibition zones arranged in a cross pattern. The zones in quadrants I, II, and III are well-defined and show significant bacterial growth inhibition.

Replikasi	Zona Bening
1	
2	
3	

Replikasi	Zona Bening
1	
2	
3	

**Lampiran 12. Hasil Pengukuran Diameter Zona Bening Amilase terhadap Suhu**

Karakterisasi suhu 30°C

<b>Bakteri</b>	<b>Diameter Zona Bening (mm)</b>			<b>Rata-rata (mm)</b>	<b>SD</b>
	<b>Replikasi 1</b>	<b>Replikasi 2</b>	<b>Replikasi 3</b>		
<i>E. coli</i>	19,65	21,98	24,60	22,08	2.48
<i>P. aeruginosa</i>	12,38	20,08	18,38	16,95	4.06
<i>S. typhi</i>	17,63	12,98	19,93	16,85	3.54
<i>S. marcescens</i>	13,48	13,18	23,23	16,63	5.72
<i>S. aureus</i>	12,25	12,13	17,33	13,90	2.97

Karakterisasi suhu 40°C

<b>Bakteri</b>	<b>Diameter Zona Bening (mm)</b>			<b>Rata-rata (mm)</b>	<b>SD</b>
	<b>Replikasi 1</b>	<b>Replikasi 2</b>	<b>Replikasi 3</b>		
<i>E. coli</i>	19,03	17,55	25,38	20,65	4,16
<i>P. aeruginosa</i>	15,58	14,78	15,78	15,38	0,53
<i>S. typhi</i>	17,45	19,15	18,58	18,39	0,87
<i>S. marcescens</i>	12,35	14,90	20,88	16,04	4,38
<i>S. aureus</i>	11,20	17,15	19,45	15,93	4,23

Karakterisasi suhu 50°C

<b>Bakteri</b>	<b>Diameter Zona Bening (mm)</b>			<b>Rata-rata (mm)</b>	<b>SD</b>
	<b>Replikasi 1</b>	<b>Replikasi 2</b>	<b>Replikasi 3</b>		
<i>E. coli</i>	18,55	18,68	20,18	19,14	0,91
<i>P. aeruginosa</i>	17,85	18,00	15,90	17,25	1,17
<i>S. typhi</i>	17,65	22,56	15,63	18,61	3,56
<i>S. marcescens</i>	13,85	26,88	19,10	19,94	6,56
<i>S. aureus</i>	20,38	17,95	16,45	18,26	1,98

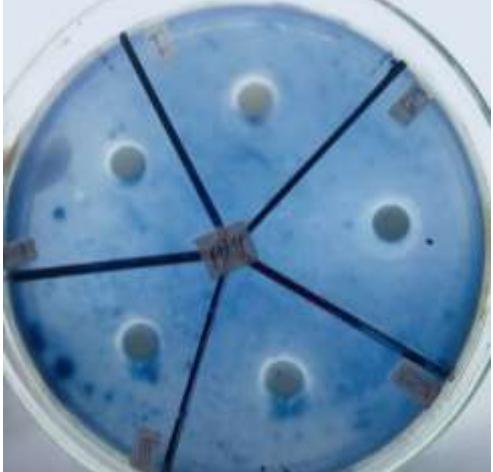
Karakterisasi suhu 60°C

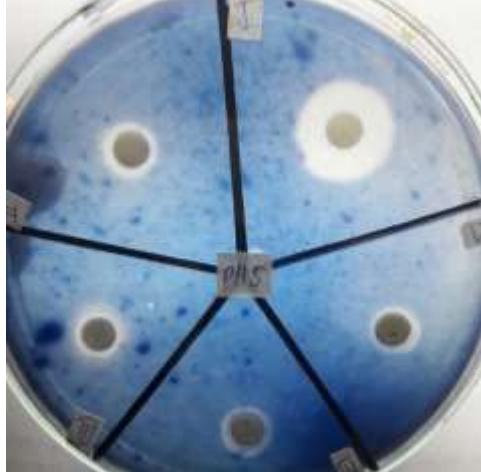
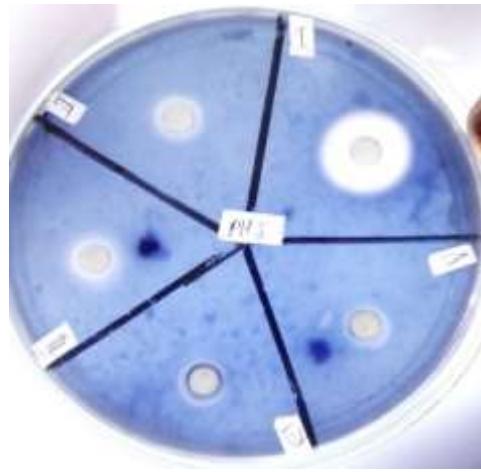
<b>Bakteri</b>	<b>Diameter Zona Bening (mm)</b>			<b>Rata-rata (mm)</b>	<b>SD</b>
	<b>Replikasi 1</b>	<b>Replikasi 2</b>	<b>Replikasi 3</b>		
<i>E. coli</i>	19,23	16,35	18,30	17,96	1,47
<i>P. aeruginosa</i>	14,25	14,20	13,08	13,84	0,66
<i>S. typhi</i>	14,40	13,88	13,78	14,02	0,33
<i>S. marcescens</i>	12,33	12,63	12,15	12,37	0,24
<i>S. aureus</i>	17,68	15,55	15,43	16,22	1,27

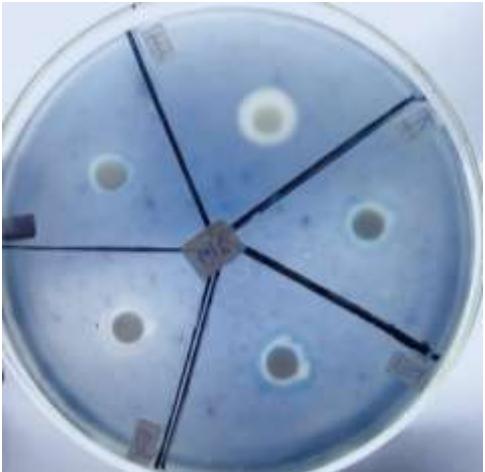
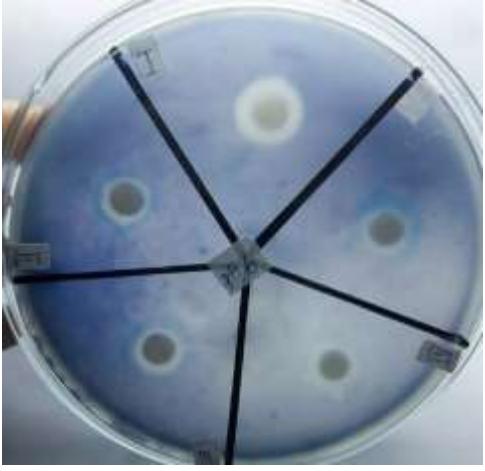
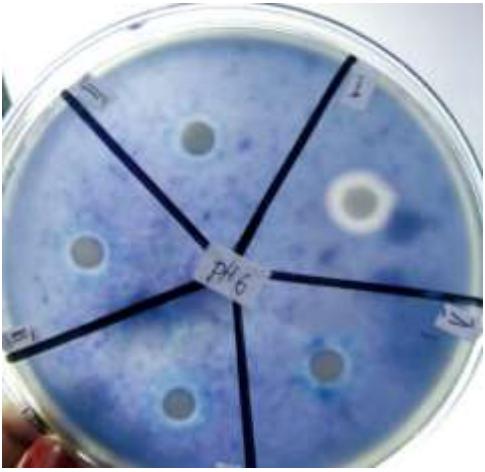
Karakterisasi suhu 70°C

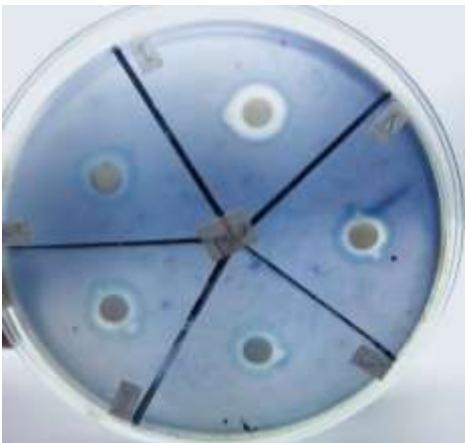
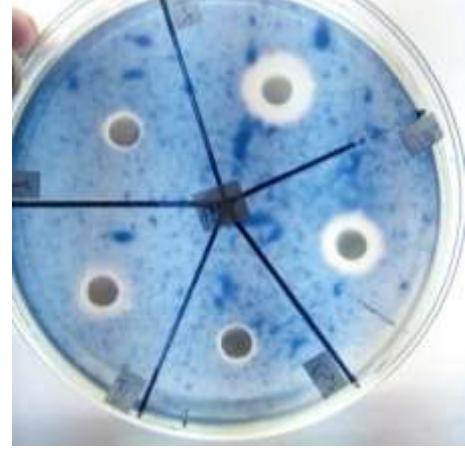
<b>Bakteri</b>	<b>Diameter Zona Bening (mm)</b>			<b>Rata-rata (mm)</b>	<b>SD</b>
	<b>Replikasi 1</b>	<b>Replikasi 2</b>	<b>Replikasi 3</b>		
<i>E. coli</i>	22,08	19,75	19,25	20,33	1,51
<i>P. aeruginosa</i>	12,55	14,43	14,33	13,77	1,06
<i>S. typhi</i>	11,53	19,05	16,15	15,58	3,79
<i>S. marcescens</i>	10,33	18,60	17,30	15,41	4,45
<i>S. aureus</i>	13,95	15,25	16,88	15,36	4,47

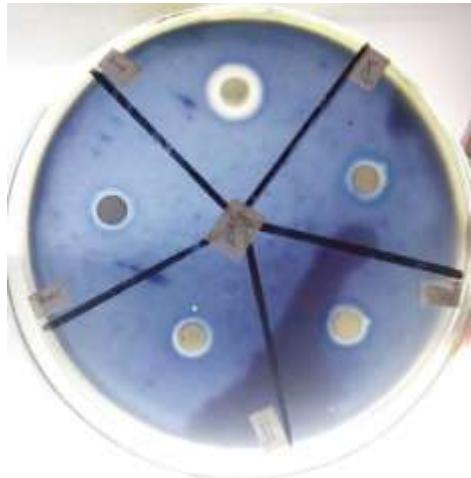
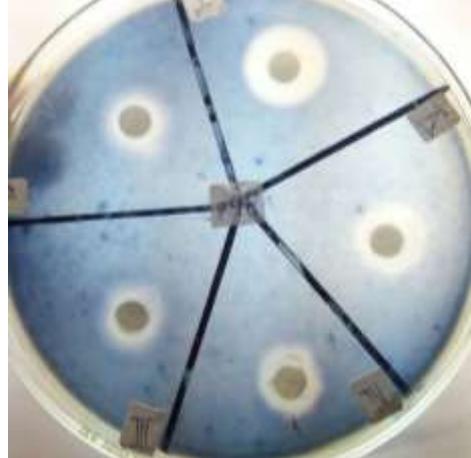
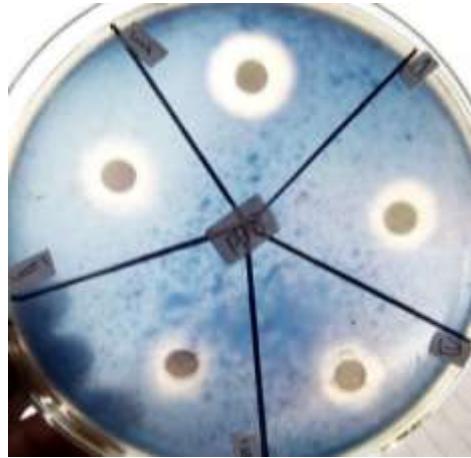
**Lampiran 13. Zona Bening Aktivitas Amilase terhadap pH**

Replikasi	Zona Bening
1	
2	

Replikasi	Zona Bening
1	
2	
3	

Replikasi	Zona Bening
1	
2	
3	

Replikasi	Zona Bening
1	
2	
3	

Replikasi	Zona Bening
1	
2	
3	

**Lampiran 14. Hasil Pengukuran Diameter Zona Bening Amilase terhadap pH**

Karakterisasi pH 4

<b>Bakteri</b>	<b>Diameter Zona Bening (mm)</b>			<b>Rata-rata (mm)</b>	<b>SD</b>
	<b>Replikasi 1</b>	<b>Replikasi 2</b>	<b>Replikasi 3</b>		
<i>E. coli</i>	9,93	8,90	9,20	9,34	0,53
<i>P. aeruginosa</i>	0	0	0	0	0
<i>S. typhi</i>	0	0	0	0	0
<i>S. marcescens</i>	0	0	0	0	0
<i>S. aureus</i>	0	0	0	0	0

Karakterisasi pH 5

<b>Bakteri</b>	<b>Diameter Zona Bening (mm)</b>			<b>Rata-rata (mm)</b>	<b>SD</b>
	<b>Replikasi 1</b>	<b>Replikasi 2</b>	<b>Replikasi 3</b>		
<i>E. coli</i>	16,18	15,53	16,68	16,13	0,58
<i>P. aeruginosa</i>	8,70	9,13	9,38	9,07	0,34
<i>S. typhi</i>	8,80	8,80	10,38	9,33	0,91
<i>S. marcescens</i>	9,23	9,58	9,90	9,57	0,34
<i>S. aureus</i>	8,73	8,95	10,18	9,29	0,78

Karakterisasi pH 6

<b>Bakteri</b>	<b>Diameter Zona Bening (mm)</b>			<b>Rata-rata (mm)</b>	<b>SD</b>
	<b>Replikasi 1</b>	<b>Replikasi 2</b>	<b>Replikasi 3</b>		
<i>E. coli</i>	13,90	13,20	12,83	13,31	0,54
<i>P. aeruginosa</i>	9,83	10,13	9,75	9,90	0,20
<i>S. typhi</i>	8,75	10,95	9,60	9,77	1,11
<i>S. marcescens</i>	10,45	10,00	9,60	10,02	0,43
<i>S. aureus</i>	10,25	10,38	8,98	9,87	0,77

Karakterisasi pH 7

<b>Bakteri</b>	<b>Diameter Zona Bening (mm)</b>			<b>Rata-rata (mm)</b>	<b>SD</b>
	<b>Replikasi 1</b>	<b>Replikasi 2</b>	<b>Replikasi 3</b>		
<i>E. coli</i>	13,85	15,25	15,58	14,89	0,92
<i>P. aeruginosa</i>	12,48	9,38	8,23	10,03	2,20
<i>S. typhi</i>	10,50	11,00	10,18	10,56	0,41
<i>S. marcescens</i>	9,95	9,05	8,95	9,32	0,55
<i>S. aureus</i>	10,18	13,15	12,85	12,06	1,64

### Karakterisasi pH 8

<b>Bakteri</b>	<b>Diameter Zona Bening (mm)</b>			<b>Rata-rata (mm)</b>	<b>SD</b>
	<b>Replikasi 1</b>	<b>Replikasi 2</b>	<b>Replikasi 3</b>		
<i>E. coli</i>	15,90	16,55	16,90	16,45	0,51
<i>P. aeruginosa</i>	9,20	13,03	14,05	12,09	2,56
<i>S. typhi</i>	8,78	12,13	12,30	11,07	1,99
<i>S. marcescens</i>	9,80	12,68	12,23	11,57	1,55
<i>S. aureus</i>	9,95	13,95	13,20	12,37	2,13

## Lampiran 15. Hasil Analisis Data Karakterisasi Suhu dengan SPSS

### Uji kolmogorov Smirnov

Tujuan : mengetahui kenormalan data sebagai syarat uji ANOVA

Kriteria uji : Sig. < 0,05 berarti H0 ditolak    Sig. > 0,05 H0 diterima

Hasil:

**One-Sample Kolmogorov-Smirnov Test**

		Aktivitas Amilase Suhu
N		75
Normal Parameters <sup>a,b</sup>	Mean	16.7857
	Std. Deviation	3.49584
	Absolute	.070
Most Extreme Differences	Positive	.070
	Negative	-.051
Kolmogorov-Smirnov Z		.605
Asymp. Sig. (2-tailed)		.858

a. Test distribution is Normal.

b. Calculated from data.

Kesimpulan : sig. 0,858 > 0,05 (H0 diterima) maka data diameter zona bening (aktivitas amilase) terdistribusi normal.

### Uji Levene

Tujuan : untuk mengetahui homogenitas

Kriteria uji : Sig. <0,05 berarti H0 ditolak. Sig. >0,05 H0 diterima

Hasil :

**Levene's Test of Equality of Error Variances<sup>a</sup>**

Dependent Variable: Aktivitas Amilase Suhu

F	df1	df2	Sig.
2.968	24	50	.001

Tests the null hypothesis that the error variance of the dependent variable is equal across groups.

a. Design: Intercept + Bakteri + Suhu + Bakteri \* Suhu

Kesimpulan :  $\text{Sig. } 0,001 < 0,05$  ( $H_0$  ditolak) maka masing-masing kelompok data aktivitas amilase berasal dari populasi dengan varians yang berbeda (tidak homogen).

### **Uji Two Way Anova**

#### **Tests of Between-Subjects Effects**

Dependent Variable: Aktivitas Amilase Suhu

Source	Type III Sum of Squares	df	Mean Square	F	Sig.
Corrected Model	444.534 <sup>a</sup>	24	18.522	2.014	.019
Intercept	21132.063	1	21132.063	2297.888	.000
Suhu	126.588	4	31.647	3.441	.015
Bakteri	210.583	4	52.646	5.725	.001
Suhu * Bakteri	107.362	16	6.710	.730	.751
Error	459.815	50	9.196		
Total	22036.412	75			
Corrected Total	904.348	74			

a. R Squared = .492 (Adjusted R Squared = .247)

Kesimpulan :

1. Faktor pertama bakteri (amilase) : karena signifikansi nilainya  $0,015 < 0,05$  ( $H_0$  ditolak), artinya dari ke lima bakteri tersebut memiliki aktivitas amilase yang tidak sama.
2. Faktor kedua suhu (amilase) : karena signifikansi nilainya  $0,001 < 0,05$  ( $H_0$  ditolak), artinya pengaruh suhu terhadap aktivitas amilase dari ke lima bakteri tersebut tidak sama.
3. Faktor interaksi/ hubungan (aktivitas amilase dengan pemberian variasi suhu) : karena signifikansi nilainya adalah  $0,751 > 0,05$  ( $H_0$  diterima), artinya amilase yang dihasilkan oleh ke lima bakteri tersebut memiliki interaksi dengan pemberian pengaruh suhu.

**Aktivitas Amilase Suhu**Tukey HSD<sup>a,b</sup>

Suhu	N	Subset	
		1	2
60oC	15	14.8827	
70oC	15	15.8440	15.8440
40oC	15	17.2807	17.2807
30oC	15	17.2807	17.2807
50oC	15		18.6407
Sig.		.210	.101

Means for groups in homogeneous subsets  
are displayed.

Based on observed means.

The error term is Mean Square(Error) =

9.196.

a. Uses Harmonic Mean Sample Size =  
15.000.

b. Alpha = .05.

Kesimpulan : Menunjukkan bahwa pada suhu 30, 40, dan 70°C tidak memiliki perbedaan yang signifikan karena berada dalam satu subset, sedangkan pada suhu 50 dan 60°C memiliki perbedaan yang signifikan karena tidak berada dalam satu subset yang sama.

## Lampiran 16. Hasil Analisis Data Karakterisasi pH dengan SPSS

### Uji kolmogorov Smirnov

Tujuan : mengetahui kenormalan data sebagai syarat uji ANOVA

Kriteria uji : Sig. < 0,05 berarti H0 ditolak    Sig. > 0,05 H0 diterima

Hasil:

One-Sample Kolmogorov-Smirnov Test

		Aktivitas Amilase pH
N		75
Normal Parameters <sup>a,b</sup>	Mean	9.4401
	Std. Deviation	4.71396
	Absolute	.264
Most Extreme Differences	Positive	.137
	Negative	-.264
Kolmogorov-Smirnov Z		2.289
Asymp. Sig. (2-tailed)		.000

a. Test distribution is Normal.

b. Calculated from data.

Kesimpulan : Sig. 0,000 > 0,05 (H0 ditolak) maka data diameter zona bening (aktivitas amilase) tidak terdistribusi normal.

### Uji Kruskal Wallis

Tujuan : mengetahui perbedaan data aktivitas amilase berdasarkan variasi pH

Kriteria uji : Sig. < 0,05 berarti H0 ditolak    Sig. > 0,05 H0 diterima

Hasil :

Test Statistics<sup>a,b</sup>

		Aktivitas Amilase pH
Chi-Square		34.551
df		4
Asymp. Sig.		.000

a. Kruskal Wallis Test

b. Grouping Variable: pH

Kesimpulan : Menunjukkan nilai sig 0,000 < 0,05 (H0 ditolak) artinya pada hasil data aktivitas amilase pH terdapat perbedaan antara pH 4-8.

**Lampiran 17. Alat Penelitian**

Inkubator



Oven



Autoclav



Jangka Sorong



Waterbath

### Lampiran 18. Cara Pembuatan Media

#### *Sulfide Indol Motility (SIM)*

Komposisi :	Pepton from casein	20	gram
	Pepton from meat	6	gram
	Ammonium Iron (II) citrate	0,2	gram
	Sodium thiosulfate	0,2	gram
	Agar-agar	0,2	gram

Cara pembuatan : Dilarutkan media SIM dalam aquadest sebanyak 1 L, dipanaskan sampai larut sempurna, kemudian disterilkan dengan autoklaf pada suhu 121°C selama 15 menit dan dituangkan dalam cawan petri.

#### *Kligler Iron Agar (KIA)*

Komposisi :	Pepton from casein	15	gram
	Pepton from meat	5	gram
	Ammonium Iron (II) citrate	0,5	gram
	Meat extract	3	gram
	Yeast extract	3	gram
	Sodium chloride	5	gram
	Laktosa	10	gram
	Glukosa	1	gram
	Sodium thiosulfate	0,5	gram
	Phenol red	0,024	gram
	Agar-agar	12	gram

Cara Pembuatan : Dilarutkan media KIA dalam aquadest sebanyak 1 L, dipanaskan sampai larut sempurna, kemudian disterilkan dengan autoklaf pada suhu 121°C selama 15 menit dan tuangkan dalam cawan petri.

#### *Lysine Indol Agar (LIA)*

Komposisi :	Pepton from meat	5	gram
	Yeast extract	3	gram

Glukosa	1	gram
Lysine monohidrochloride	10	gram
Sodium thiosulfate	0,04	gram
Ammonium Iron (II) citrate	0,5	gram
Bromo cresol purple	0,02	gram
Agar-agar	12,5	gram

Cara Pembuatan : Dilarutkan dalam aquadest sebanyak 1 L, dipanaskan sampai larut sempurna, kemudian disterilkan dengan autoklaf pada suhu 121°C selama 15 menit dan tuangkan dalam cawan petri.

### ***Simmon Citrate Agar (SCA)***

Komposisi :	Ammonium hydrogen fosfat	1	gram
	DI-Potassium hydrogen fosfat	1	gram
	Sodium chloride	5	gram
	Magnesium sulfate	0,2	gram
	Bromo thymol blue	0,08	gram
	Agar-agar	12,5	gram

Cara Pembuatan: Dilarutkan media SCA dalam aquadest sebanyak 1 L, dipanaskan sampai larut sempurna, kemudian disterilkan dengan autoklaf pada suhu 121°C selama 15 menit dan tuangkan dalam cawan petri.

### ***Nutrient Agar (NA)***

Komposisi :	Peptone	5	gram
	Sodium chloride	5	gram
	HM peptone B	1,5	gram
	Yeast extract	1,5	gram
	Agar	15	gram

Cara Pembuatan : Dilarutkan 28 gram media NA dalam 1 L aquadest. Dipanaskan sampai mendidih dan melarut sempurna. Disterilkan dengan autoklaf pada suhu 121°C selama 15 menit, lalu tambahkan 10 ml Kalium Tellurit 1 %. Kemudian dituang ke dalam cawan petri steril.

### ***Endo Agar (EA)***

Komposisi :	Peptone	10	gram
	Lactose	10	gram
	Dipotassium phosphate	3,5	gram
	Sodium sulphite	2,5	gram
	Basic fuchsin	0,5	gram
	Agar	15	gram

Cara Pembuatan : Dilarutkan 41,5 gram media EA dalam 1 L aquadest, lalu ditambahkan 1 ml sodium sulfit 10%. Dipanaskan sampai mendidih dan melarut sempurna. Disterilkan dengan autoklaf pada suhu 121°C selama 15 menit, kemudian dituang ke dalam cawan petri steril.

### ***Pseudomonas Selective Agar (PSA)***

Komposisi :	Pancreatic digest of gelatin	20	gram
	Magnesium chloride	1,4	gram
	Dipotassium sulphate	10	gram
	Cetrimide	0,3	gram
	Agar	13,6	gram

Cara Pembuatan : Dilarutkan 45,3 gram media PSA dalam 1 L aquadest, lalu tambahkan 10 ml gliserol. Dipanaskan sampai mendidih dan melarut sempurna. Disterilkan dengan autoklaf pada suhu 121°C selama 15 menit, kemudian dituang ke dalam cawan petri steril.

### ***Salmonella-Shigella Agar (SSA)***

Komposisi :	Proteose peptone	5	gram
	Lactose	10	gram
	Bile salts mixture	8,5	gram
	Sodium citrate	8,5	gram
	Sodium thiosulphate	8,5	gram
	Ferric citrate	1	gram

Brilliant green	0,0003 gram
Neutral red	0,025 gram
Agar	13,5 gram

Cara Pembuatan : Dilarutkan 60 gram media PSA dalam 1 L aquadest Dipanaskan sampai mendidih dan molarut sempurna. Disterilkan dengan autoklaf pada suhu 121°C selama 15 menit, kemudian dituang ke dalam cawan petri steril.

#### ***Vogel Johnson Agar (VJA)***

Komposisi :	Tryptone	10	gram
	Yeast extract	5	gram
	Mannitol	10	gram
	Dibasic potassium phosphate	5	gram
	Lithium chloride	5	gram
	Glycine	10	gram
	Phenol red	0.025	gram
	Agar	16	gram

Cara Pembuatan : Dilarutkan 61 gram media PSA dalam 1 L aquadest Dipanaskan sampai mendidih dan molarut sempurna. Disterilkan dengan autoklaf pada suhu 121°C selama 15 menit, lalu tambahkan 10 ml Kalium Tellurit 1 %. Kemudian dituang ke dalam cawan petri steril.

## Lampiran 19. Cara Pembuatan Larutan Buffer

### Buffer Sitrat pH 4

Dilarutkan 0,492 g Sodium Citrate Dihydrate dalam 40 ml aquadest steril, ditambahkan 0,639 g Citric Acid ke dalam larutan dan diaduk ad homogen. Ditambahkan larutan HCl atau NaOH sampai pH yang diinginkan sesuai, kemudian tambahkan aquadest steril ad 50 ml.

### Buffer Sitrat pH 5

Dilarutkan 0,741 g Sodium Citrate Dihydrate dalam 40 ml aquadest steril, ditambahkan 0,476 g Citric Acid ke dalam larutan dan diaduk ad homogen. Ditambahkan larutan HCl atau NaOH sampai pH yang diinginkan sesuai, kemudian tambahkan aquadest steril ad 50 ml.

### Buffer Sitrat pH 6

Dilarutkan 1,204 g Sodium Citrate Dihydrate dalam 40 ml aquadest steril, ditambahkan 0,174 g Citric Acid ke dalam larutan dan diaduk ad homogen. Ditambahkan larutan HCl atau NaOH sampai pH yang diinginkan sesuai, kemudian tambahkan aquadest steril ad 50 ml.

### Buffer Fosfat pH 7

Dilarutkan 0,774 g Sodium phosphate dibasic heptahydrate dalam 40 ml aquadest steril, ditambahkan 0,291 g Sodium phosphate monobasic monohydrate ke dalam larutan dan diaduk ad homogen. Ditambahkan larutan HCl atau NaOH sampai pH yang diinginkan sesuai, kemudian tambahkan aquadest steril ad 50 ml.

### Buffer Fosfat pH 8

Dilarutkan 0,814 g Sodium phosphate dibasic heptahydrate dalam 40 ml aquadest steril, ditambahkan 0,044 g Sodium phosphate monobasic monohydrate ke dalam larutan dan diaduk ad homogen. Ditambahkan larutan HCl atau NaOH sampai pH yang diinginkan sesuai, kemudian tambahkan aquadest steril ad 50 ml.

## Lampiran 20. Hasil Clustal Omega

MSITVANSDDDBLVTAIQQRIALESTSYVVEGAYPAKTESDDIBLARIATADCHEVILGAEVWRVG MSEIDWRSSTVYQTYKEFSDIDTGIGDINGIEKDYKLIGDYIYLTVIVSPMONGDISYL MKLLPFLMLISPALASWLGEFADEIAAGFTSQALPQGQPERLYDQHCPAPAKLKNQI MKLAALFLFLIGEVAIASWSSEGFPASEQGTGIVSHAOQPGRETLINEDOCOPADAKLKNQI MKLAAPATLFLIGEATIASNTSSEGFPFLSTQGTGFSHAALTGTRALTHDQJCPSALKLNQI	ETAREELPLFENDFWSAQATTPFGCERYFRIEAMID-RFAGRRELLRAKHAR--IP ----- ----- ----- ----- ----- ----- ----- ----- ----- ----- ----- ----- ----- ----- ----- -----	56 0 37 36 36 36
MSITVANSDDDBLVTAIQQRIALESTSYVVEGAYPAKTESDDIBLARIATADCHEVILGAEVWRVG MSEIDWRSSTVYQTYKEFSDIDTGIGDINGIEKDYKLIGDYIYLTVIVSPMONGDISYL MKLLPFLMLISPALASWLGEFADEIAAGFTSQALPQGQPERLYDQHCPAPAKLKNQI MKLAALFLFLIGEVAIASWSSEGFPASEQGTGIVSHAOQPGRETLINEDOCOPADAKLKNQI MKLAAPATLFLIGEATIASNTSSEGFPFLSTQGTGFSHAALTGTRALTHDQJCPSALKLNQI	LDLIREGEDELLQCAEGPFELAC-A-PLAE--RLQ-----ACQSVERTVNLQ ----- ----- ----- ----- ----- ----- ----- ----- ----- ----- ----- ----- ----- ----- -----	107 0 97 96 95
MSITVANSDDDBLVTAIQQRIALESTSYVVEGAYPAKTESDDIBLARIATADCHEVILGAEVWRVG MSEIDWRSSTVYQTYKEFSDIDTGIGDINGIEKDYKLIGDYIYLTVIVSPMONGDISYL MKLLPFLMLISPALASWLGEFADEIAAGFTSQALPQGQPERLYDQHCPAPAKLKNQI MKLAALFLFLIGEVAIASWSSEGFPASEQGTGIVSHAOQPGRETLINEDOCOPADAKLKNQI MKLAAPATLFLIGEATIASNTSSEGFPFLSTQGTGFSHAALTGTRALTHDQJCPSALKLNQI	TGELLIRVGPREHLVRSSEYVEVERPLAESWELPRESSEDFIRHGTFFDV ----- ----- ----- ----- ----- ----- ----- ----- ----- ----- ----- ----- ----- ----- -----	163 0 149 148 147
MSITVANSDDDBLVTAIQQRIALESTSYVVEGAYPAKTESDDIBLARIATADCHEVILGAEVWRVG MSEIDWRSSTVYQTYKEFSDIDTGIGDINGIEKDYKLIGDYIYLTVIVSPMONGDISYL MKLLPFLMLISPALASWLGEFADEIAAGFTSQALPQGQPERLYDQHCPAPAKLKNQI MKLAALFLFLIGEVAIASWSSEGFPASEQGTGIVSHAOQPGRETLINEDOCOPADAKLKNQI MKLAAPATLFLIGEATIASNTSSEGFPFLSTQGTGFSHAALTGTRALTHDQJCPSALKLNQI	SSEGGILLEPATA-----KELAFSWNMVATVFLDRENGEPANDSAYSEREDGQ SSEGGILLEPAAED-----TSAPEWNVATVFLDRENGEPANDSAYSEREDGQ SSEGGILLEPAAH-----ASAPTNWNVATVFLDRENGEPANDSAYSEREDGQ ----- ----- ----- ----- -----	208 0 202 201 200
MSITVANSDDDBLVTAIQQRIALESTSYVVEGAYPAKTESDDIBLARIATADCHEVILGAEVWRVG MSEIDWRSSTVYQTYKEFSDIDTGIGDINGIEKDYKLIGDYIYLTVIVSPMONGDISYL MKLLPFLMLISPALASWLGEFADEIAAGFTSQALPQGQPERLYDQHCPAPAKLKNQI MKLAALFLFLIGEVAIASWSSEGFPASEQGTGIVSHAOQPGRETLINEDOCOPADAKLKNQI MKLAAPATLFLIGEATIASNTSSEGFPFLSTQGTGFSHAALTGTRALTHDQJCPSALKLNQI	AIGSEEGGEHEALTHPELGREDERFLIVVYAEHNEELAIIQGS-PDHMELEH----- ----- ----- YHGYYLDTIDAMNGEALRIVYSAHQGIRLUDVNNHGTATIA-----DMQT YHGYYLDTIDAMNGEALRIVYSAHQGIRLUDVNNHGTATIA-----DME YHGYYLDTIDAMNGEALRIVYSAHQGIRLUDVNNHGTATIA-----DME ----- ----- ----- -----	262 50 257 256 255

MSSIVANSDDPLVIAQQRRALESYSPEVEGAYPAKTESDIDLARIAARIADGHETVLGAEVWRRVG MSKEIDWRSVTVYQIYPSSTDITGIGTINGIEKDYKLIGVYIYLTVYESPMDNGDISYL MKLLEPLMILISPEALASWTLSEPAFDETAGIFTQALIKRGQFLRLYDHCOPAFAKIQLI MKLAACFLILEGEVAASWTSPEFAFSEQTGFYSHAQLPKGTRTLNDQOCOPADAIIKQML MKLAAPALLTIGEIAASSWTSPEFPILSTQETGFTSHALITGTRALTHDQOCOPSEAIKQML	MSSIVANSDDPLVIAQQRRALESYSPEVEGAYPAKTESDIDLARIAARIADGHETVLGAEVWRRVG MSKEIDWRSVTVYQIYPSSTDITGIGTINGIEKDYKLIGVYIYLTVYESPMDNGDISYL MKLLEPLMILISPEALASWTLSEPAFDETAGIFTQALIKRGQFLRLYDHCOPAFAKIQLI MKLAACFLILEGEVAASWTSPEFAFSEQTGFYSHAQLPKGTRTLNDQOCOPADAIIKQML MKLAAPALLTIGEIAASSWTSPEFPILSTQETGFTSHALITGTRALTHDQOCOPSEAIKQML	MSSIVANSDDPLVIAQQRRALESYSPEVEGAYPAKTESDIDLARIAARIADGHETVLGAEVWRRVG MSKEIDWRSVTVYQIYPSSTDITGIGTINGIEKDYKLIGVYIYLTVYESPMDNGDISYL MKLLEPLMILISPEALASWTLSEPAFDETAGIFTQALIKRGQFLRLYDHCOPAFAKIQLI MKLAACFLILEGEVAASWTSPEFAFSEQTGFYSHAQLPKGTRTLNDQOCOPADAIIKQML MKLAAPALLTIGEIAASSWTSPEFPILSTQETGFTSHALITGTRALTHDQOCOPSEAIKQML
-----PIGTAEPIGSTAELIP----- NPARDYFFR-----SEDPENWHSK-----EGNAYDE FHESSYLOGAVENTIGKWSNDREPGWHSFUDINEDAGREWGNWRD YQFAYLSSEDYKKSIGERSDKPAAGTHSFUDINEDAGREWGNWRD YQFAYLSSGARQITIGDRNTWPHAGWSHSDINFSLSAEMKWKMRD -----	-----PIGTAEPIGSTAELIP----- PK-----KEDTY-NVDEY-----AEALPSIN-----EALAD TDEY-----YLIHFDSQDLDNWNPYRQLR IGDYDFGDDITSLATLPKTEAGASPLTRHPDTARD--MEGATRILY IGDYDFGDDITSLATLPKTEASTASGEFVYVANWDHVA--IGITPDRYLTH IGDYDFGDDITSLATLPKTEESTTSGEFVYANKPDRYLTH--IGITPDRYLTH -----*	-----PIGTAEPIGSTAELIP----- VILGW-VEQGVLFREVNDVPHKELPENWLAEVGRHEFQVTEAETHPAMALGV IVHNW-DIGVNGRFDTUNISGEYKODKG----- WLSQWHDGIGFVDAHVEPTALKQA----- WLSQWHDGIGFVDAHVEPANQKXEA----- WLSQWHDGIGFVDAHVEPANQKXOA----- -----* : * : ** : * : -----
-----GFSQSTYFTRNDQELAEYTAELQPFRCYRPNFTVNTPDINWELQSGRGFLI -----KEFTIGPRVHEFLILRQ-TFG----- -----I-----T-----ALLAWZEHHQ-ALD----- -----S-----AHLREKTAENDK-ALD----- -----S-----AHLHEWQANDK-ALD----- -----* : * : -----	-----GFSQSTYFTRNDQELAEYTAELQPFRCYRPNFTVNTPDINWELQSGRGFLI -----KEFTIGPRVHEFLILRQ-TFG----- -----I-----T-----ALLAWZEHHQ-ALD----- -----S-----AHLREKTAENDK-ALD----- -----S-----AHLHEWQANDK-ALD----- -----* : * : -----	-----RAIAIAMSGLGMYSGELCEAALFGE----- -----NTDMTGEN-----SITIENCIRKTQPERELNSTNFHJKVYDGEKNTAKDF----- -----DAP-----TMIGEANGGNGKSDY -----DKP-----TMIGEANGGNGQSDY -----DSP-----TMIGEANGGNGKSDY -----* : * : -----



## Lampiran 21. Phylogenetic Tree

